

CrossMark
click for updatesCite this: *RSC Adv.*, 2015, 5, 54102

A high performance electrochemical biosensor based on Cu₂O–carbon dots for selective and sensitive determination of dopamine in human serum

Qitong Huang,^{*a} Xiaofeng Lin,^{*c} Changqing Lin,^a Yong Zhang,^{ab} Shirong Hu^c and Chan Wei^c

A green and facile method was developed by synthesizing a cuprous oxide–carbon dots/Nafion (Cu₂O–CDs/NF) composite film for highly sensitive and reliable determination of dopamine (DA). The Cu₂O nanoparticles could improve the conductivity of the electrode, while the CDs with carboxyl groups and the NF with sulfo groups could attract cations *via* the ion-exchange model and exclude anions by the electrostatic action. The proposed biosensor exhibited a low detection limit of 1.1 nM with a wide linear range of 0.05–45.0 μM and acquired excellent sensitivity and selectivity for DA. Furthermore, the Cu₂O–CDs/NF/GCE also was applied to determine DA in human serum with satisfactory results and showed good activity for more than two months.

Received 27th March 2015

Accepted 12th June 2015

DOI: 10.1039/c5ra05433h

www.rsc.org/advances

1. Introduction

In recent years, nanomaterials have made an important impact on diverse science, engineering, and commercial sectors due to their high catalytic activity, low cost, and good stability.^{1–5} Cuprous oxide (Cu₂O), as a p-type semiconductor, has drawn intense interest in the past decade, not only for its unique properties but also for its applications in various fields such as gas sensors,⁶ solar cells,⁷ lithium ion batteries,⁸ catalysis⁹ and so on. As a star type of material, Cu₂O nanomaterials have been continually used as an ideal material for fabricating electrochemical biosensors due to their ultrahigh electrocatalysis capacity and surface area effect in biosensing analysis.¹⁰ Due to the ‘zero-dimensional’ feature, carbon dots (CDs) had become one of the hottest materials in many fields, such as physics, chemistry, biological imaging and electronics since their discovery in 2004.^{11–13} The advantages of low toxicity, excellent biocompatibility, high chemical stability and remarkable conductivity, which make it as a good electrode material to construct various electrochemical platforms.¹⁴ What's more, the CDs also can be used as reducing agents,^{15,16} which can directly reduce copper hydroxide to form Cu₂O–CDs without adding other reducers or stabilizers.

Belonging to the family of excitatory chemical neurotransmitters, dopamine (DA) acts as one of the most significant

catecholamines, which plays a pivotal role in the function of human metabolism, cardiovascular, renal, central nervous and hormonal systems.^{17–20} As known, electrochemical biosensors have attracted tremendous efforts recently as a novel analytical tool due to its selectivity, sensitivity, simplicity and rapid response time.^{21–24} Due to DA's electrochemical activity, DA detection is always attracting the intense interest in electroanalysis, such as gold nanoparticles–reduced graphene oxide sheets–indium tin oxide-coated glass (AuNPs–rGOS–ITO),¹⁷ copper nanoparticles–multi-walled carbon nano-tubes/glassy carbon electrode (Cu–MWCNT/GCE),¹⁸ WO₃ nanoparticles,¹⁹ and so on. In this work, a green and economical composite of Cu₂O–CDs/Nafion (NF) composite was utilized to construct a novel dopamine (DA) biosensor. To the best of our knowledge, electrochemical detection of DA utilizing the Cu₂O–CDs/NF based hybrid film has not been published. The Cu₂O nanoparticle could increase surface area of the GCE, the CDs had carboxyl groups and the NF had sulfo groups, they could attract cations *via* the ion-exchange model and exclude anions by the electrostatic action, which could improve the selectivity in the detection of DA.^{16,25} Therefore, the Cu₂O–CDs/NF composite provided an excellent platform for sensitive, selective and reliable determination of DA. In addition, the modified electrode was applied to detect DA content in the human serum with satisfactory results.

2. Experimental

2.1. Reagents and instrumentation

Glucose, ethanol, ascorbic acid, uric acid and dopamine were purchased from Sinopharm Chemical Reagent Co., Ltd.

^aDepartment of Food and Biological Engineering, Zhangzhou Institute of Technology, Zhangzhou, 363000, PR China. E-mail: hqtblue@163.com; Fax: +86 596 2660090; Tel: +86 596 2660090

^bResearch of Environmental Science, Xiamen University, Xiamen, 361005, PR China

^cCollege of Chemistry and Environment, Minnan Normal University, Zhangzhou, 363000, PR China. E-mail: Lxf0596@163.com

(Shanghai, China). Sodium hydroxide, oleic acid and copper nitrate were purchased from Xilong Chemical Co., Ltd. (Guangdong, China). Nafion was purchased from Sigma-Aldrich Chem. Co. (USA) (5 wt% in mixture of lower aliphatic alcohols and H₂O). All the solutions were prepared with deionized water (18.2 MΩ).

All of electrochemical experiments were performed with CHI 660E electrochemical workstation (Shanghai Chenhua Instruments Co., China). A conventional three-electrode system was used in all electrochemical experiments, which consist a platinum wire auxiliary electrode, a Ag|AgCl (saturated KCl) reference electrode and a bare or modified GCE as working electrode, which. Scanning electron microscope (SEM) imaging was performed on a S-4800 electron microscope (Hitachi, Ltd., Japan). TEM images were measured using a JEM-100CX electron microscope from JEOL Ltd (Aichi Kariya, Japan). The surface morphology of the Cu₂O–CDs/NF film was characterized using atomic force microscopy (AFM, CSPM5500, China). All experiments were carried out at room temperature.

2.2. Synthesis of Cu₂O–CDs composite and Cu₂O–CDs/NF composite film

The synthesis of CDs were synthesized follow previous report.²⁶ Cu₂O–CDs composite were synthesized by adding 100 μL aqueous solution of Cu(NO₃)₂ (1.0 mol L⁻¹) was added to 200 μL aqueous solution of NaOH (1.0 mol L⁻¹). After 20 min, 100 μL CDs solution (8.0 mg mL⁻¹) was added to the above solution. Then the mixed solution was kept at 90 °C for 30 min to yield a stable purple solution of Cu₂O–CDs. After that, the Cu₂O–CDs/NF composite film was carried out: 50.0 μL NF was added in 450.0 μL ethanol to forming homogeneous solution. Then 100.0 μL Cu₂O–CDs solution was added in Nafion ethanol solution with vigorous ultrasonication.

3. Results and discussion

3.1. Characterization of CDs, Cu₂O–CDs and Cu₂O–CDs/NF/GCE

The TEM image of the CDs was shown in the Fig. 1A. It can be seen that the sizes of CDs were ranging from 1.0 nm to 5.0 nm. The Cu₂O–CDs (Fig. 1B) was characterized by SEM with the sizes ranging from 30.0 nm to 70.0 nm. The UV-vis spectrum of CDs and Cu₂O–CDs were shown in the Fig. 1C, the CDs had a strong absorption band appear at 281 nm and. As known the absorption band of Cu₂O nanocubes (<100 nm) was range from 450 nm to 490 nm.²⁷ However, the Cu₂O–CDs exhibited two characteristic absorption peaks at 284 nm and 426 nm, due to the CDs' impact, the peak of Cu₂O had a blue-shift. The XRD (Fig. 1D) was also showed the nanocompound of Cu₂O–CDs were formed. The XRD data of the three as-synthesized samples were all in good agreement with those of Cu₂O (JCPDS no. 65-3288), the 5 typical peaks located at 29.70°, 36.41°, 42.25°, 61.27° and 73.54°, which were attributed to the (110), (111), (200), (220) and (311) planes of cuprous oxide, respectively. No characteristic peaks arising from Cu or CuO could be observed in the XRD patterns, indicating that the products obtained *via* our synthetic

routes consist of only Cu₂O phase. In addition, a broad peak located at 20.02° which indicated the CDs existed in the nanocompound.^{9,26} Furthermore, 3D AFM images of the bare GCE, NF/GCE, CDs/NF/GCE and Cu₂O–CDs/NF/GCE were shown in Fig. 2. Fig. 2A showed that the bare GCE was relatively flat and smooth with the average roughness of +1.22 nm, while after NF (Fig. 2B) and CDs/NF (Fig. 2C) modification, the average roughness increased to +1.73 nm and +2.51 nm, respectively. However, for Cu₂O–CDs/NF/GCE, an average roughness of +10.42 nm was observed (Fig. 2D). From the AFM results, it was also obtained that the largest height of Cu₂O–CDs/NF/GCE was determined to be 48.27 nm, which is obviously larger than the bare GCE (5.25 nm), NF (8.03 nm) and CDs/NF (12.91 nm). These results demonstrated that Cu₂O–CDs/NF had been anchored on GCE and improved the surface effect of the electrode interface.

3.2. Cyclic voltammetric behavior of DA on the Cu₂O–CDs/NF/GCE

Fig. 3 showed the typical cyclic voltammograms (CVs) of DA on the bare GCE (a), NF/GCE (b) CDs/NF/GCE (c) and Cu₂O–CDs/NF/GCE in 0.1 mol L⁻¹ pH 7.0 PBS at the scan rate of 0.1 V s⁻¹. Well resolved, defined and more enhanced anode current peak is observed on Cu₂O–CDs/NF/GCE compared to GCE, NF/GCE, CDs/NF/GCE. The oxidation peak current intensity at Cu₂O–CDs/NF/GCE was increased 3.16 times, 2.08 times, 1.88 times higher than those obtained at GCE, NF/GCE and CDs/NF/GCE, respectively. A possible reaction mechanism of Cu₂O–CDs/NF/GCE with DA was discussed that the Cu₂O nanoparticle could make the surface of the electrode more conductive, the CDs had carboxyl groups and NF had sulfo groups which could make it have good stability for sensitive and selective determination of DA.

3.3. Effects of pH and scan rate on DA detection at Cu₂O–CDs/NF/GCE

The effect of the pH of PBS on dopamine detection at Cu₂O–CDs/NF/GCE was examined by CVs, and the results were shown in Fig. 4A. The results showed that the oxidation peak currents of DA first increased as the pH increase from 5.0 to 7.0, and then decreased as pH further increased from 7.0 to 9.0. The oxidation peak current reached a maximum at pH = 7.0. Therefore, the pH = 7.0 was used in the experiments to achieve the highest sensitivity. At the same time, the oxidation peak potential shifted negatively with the increase of pH value, indicating that protons involved in the electrode reaction. A good linear relationship between E_{pa} and pH was established by fitting with a linear regression equation as E_{pa} (V) = -0.0574pH + 0.722 ($R^2 = -0.997$, $n = 5$). The slope value of -57.4 mV per pH showed that the electron transfer was accompanied by an equal number of protons.²⁸

In order to further investigate the electrochemical behavior of DA at the Cu₂O–CDs/NF/GCE, Fig. 4B presents CVs of DA in 0.1 M PBS (pH 7.0) at the Cu₂O–CDs/NF/GCE at different scan rate. The redox peak currents increases monotonically as scan rate increases. Both of the anodic and cathodic peak currents

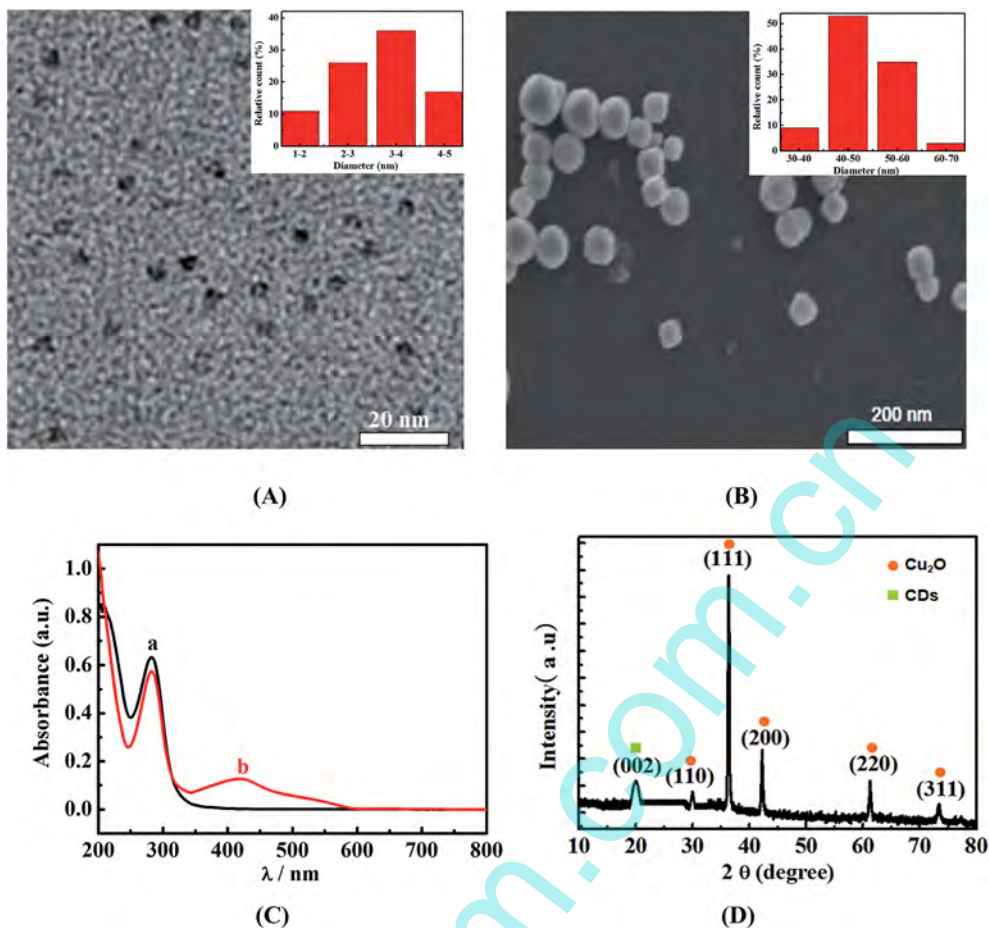


Fig. 1 (A) TEM image of CDs (B) SEM image of Cu_2O -CDs, (C) the UV-vis spectra for CDs (a) and Cu_2O -CDs composites (b), (D) XRD pattern for the obtained Cu_2O -CDs.

follow a linear relationship with the scan rate in a range from 0.05 to 0.5 V s^{-1} (Fig. 4C), for anodic peak currents was $I_{\text{pa}} (\mu\text{A}) = -12.124v - 2.943$ ($R^2 = 0.997$, $n = 5$). For cathodic peak current was $I_{\text{pc}} (\mu\text{A}) = 7.983v - 0.771$ ($R^2 = 0.995$, $n = 5$). This implied that the electrochemical oxidation of DA at the

Cu_2O -CDs/Nafion/GCE was a surface-controlled process, but not a diffusion-controlled process. In addition, the influence of scan rate on the peak potential was also investigated by CVs. As shown in the Fig. 4D, the anodic and cathodic peak currents peak potential followed a linear relationship with the logarithm

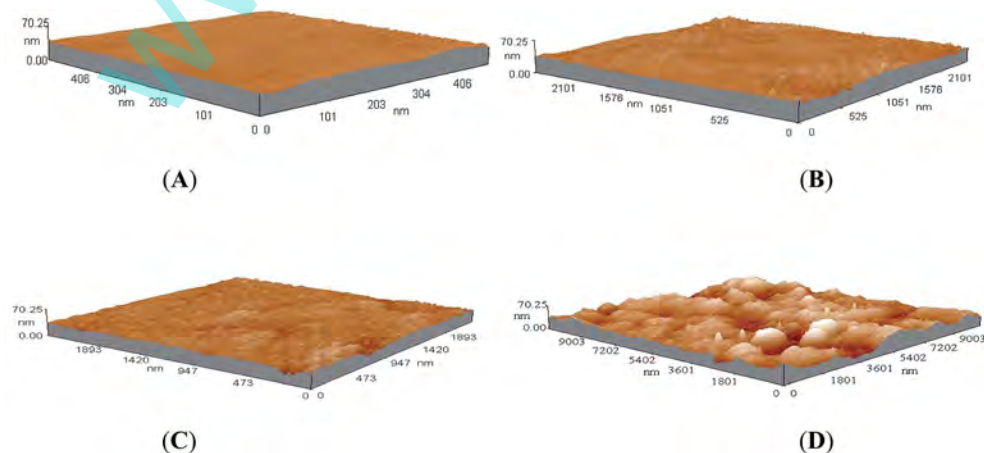


Fig. 2 3D AFM images of bare GCE (A), NF/GCE (B), CDs/NF/GCE (C) and Cu_2O -CDs/NF/GCE (D).

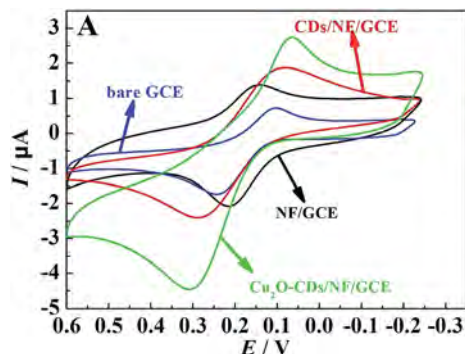


Fig. 3 CVs of 0.1 mM DA recorded on bare GCE, NF/GCE, CDs/NF/GCE and Cu₂O-CDs/NF/GCE in the pH 7.0 phosphate buffer solution (scan rate: 0.1 V s⁻¹).

of scan rate. The regression equations were E_{pa} (V) = 0.0442 log ν + 0.357 ($R^2 = 0.995$, $n = 5$) and E_{pc} (V) = -0.0291 log ν + 0.0367 ($R^2 = 0.997$, $n = 5$), respectively. The relationship between the potential and scan rate could be described as following equations by Laviron.²⁹

$$E_{pa} = E^{0'} + \frac{2.3RT}{(1-\alpha)nF} \log \nu \quad (1)$$

$$E_{pc} = E^{0'} - \frac{2.3RT}{\alpha nF} \log \nu \quad (2)$$

$$\log k_s = \alpha \log(1-\alpha) + (1-\alpha) \log \alpha - \log \frac{RT}{nFv} - \frac{(1-\alpha)\alpha nF\Delta E_p}{2.3RT} \quad (3)$$

where α was the electron transfer coefficient, n was the number of transfer electron, k_s was the standard heterogeneous rate constant, R , T and F had their usual significance. After making computations: $\alpha = 0.603$, $n = 2.32$, $k_s = 1.48$ s⁻¹. The value of k_s was bigger than some reported values,²³ indicating a fast electron transfer reaction on the Cu₂O-CDs/NF/GCE.

3.4. Interference effect

As known that the oxidation peak potentials for ascorbic acid (AA), uric acid (UA) and DA were very close to each other on a bare GCE, hence, it was difficult to separate these compounds due to their overlapping signals.³⁰ Now, the problem could be eliminated in this study. Using the Cu₂O-CDs/NF/GCE as the working electrode, the CDs had carboxyl groups and the NF had sulfo groups could prevent anionic AA and UA from reaching the electrode surface, which was in its anionic form at the working electrode surface in the pH 7.0 phosphate buffer solution, both AA (acidity coefficient, pK_a = 4.10) and UA (pK_a = 5.75) were negatively charged, but DA (pK_a = 8.89) was positively charged at physiological pH = 7.0.³¹ As shown in Fig. 5, it could be concluded that the presence of AA and UA did not interfere in the DA determination.

Furthermore, other influences from common co-existing substances were also investigated. When the relative error

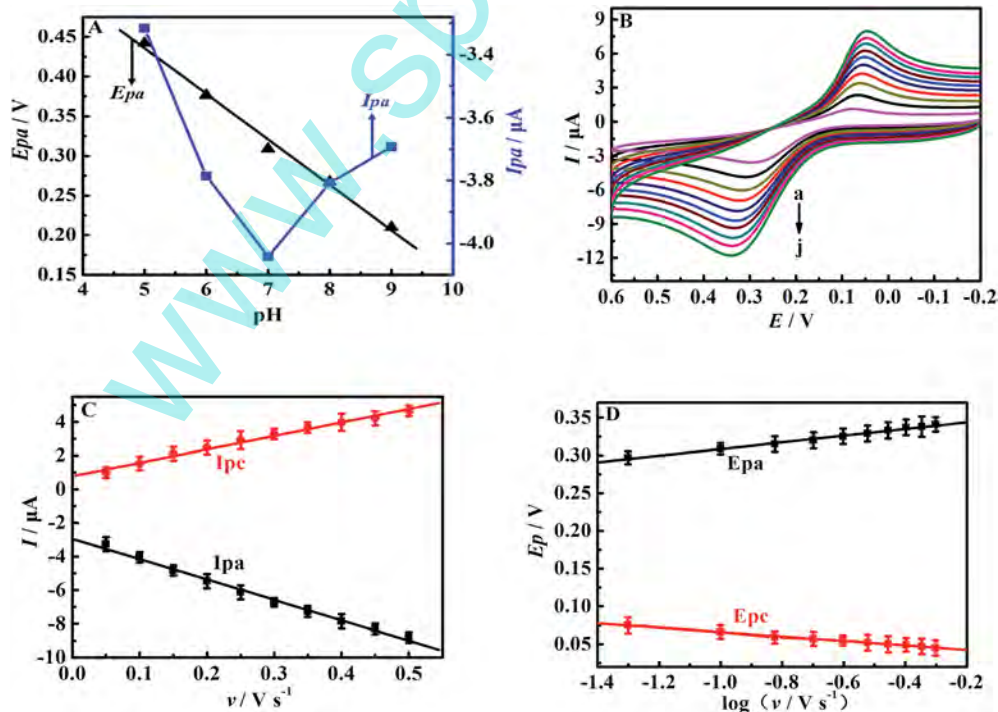


Fig. 4 (A) The relationship of the E_{pa} and I_{pa} against pH (scan rate: 0.1 V s⁻¹); (B) CV of 0.1 mM DA on Cu₂O-CDs/NF/GCE in the pH 7.0 phosphate buffer solution at various scan rates (a–j: 0.05, 0.10, 0.15, 0.20, 0.25, 0.30, 0.35, 0.40, 0.45, 0.50 V s⁻¹); (C) the relationships of I_{pa} and I_{pc} with ν (D) the relationships of E_{pa} and E_{pc} with log ν .

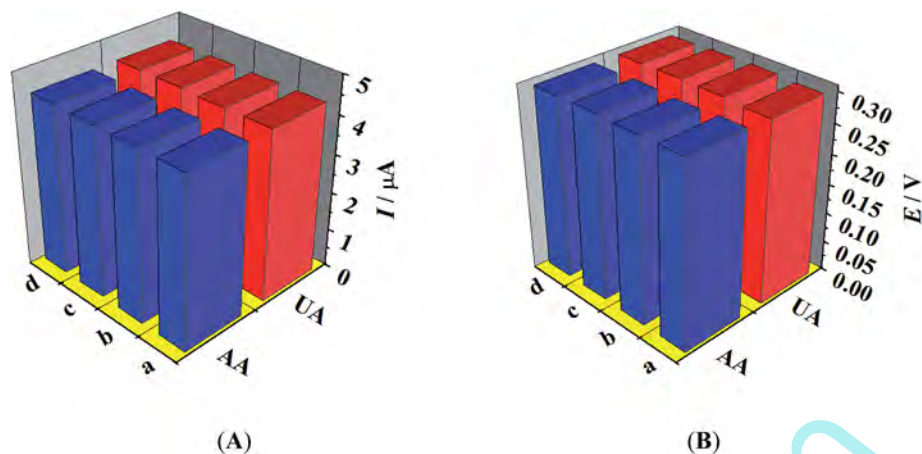


Fig. 5 The anode current peak (A) and zeta-potential (B) of 0.1 mM DA with the different concentrations of AA and UA: 0 mM (a), 10.0 mM (b), 50.0 mM (c) and 80.0 mM (d).

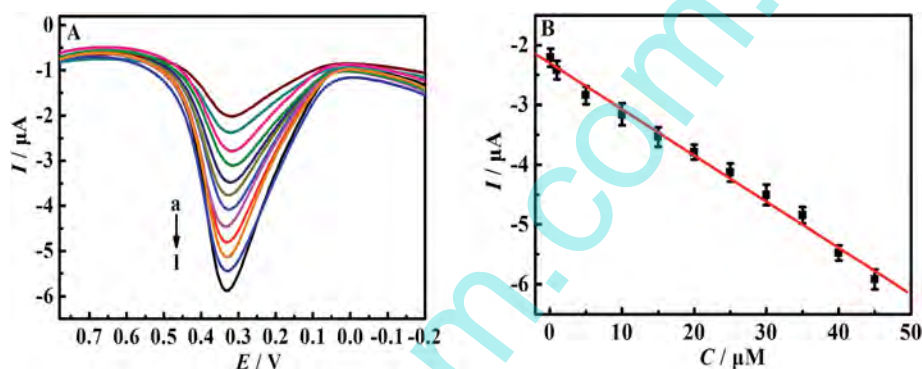


Fig. 6 (A) DPVs of DA with increasing concentration (from a to l: 0.05, 1.0, 5.0, 10.0, 15.0, 20.0, 25.0, 30.0, 35.0, 40.0, 45.0 μM). (B) The relationship of the I_{pa} with the concentration of DA.

(E_T) exceeded 5%, each matter was considered as an interfering agent. It was found that most ions and common substances at high concentration only caused negligible change: Na^+ , K^+ , NO_3^- , Cl^- , SO_4^{2-} (>200 fold), Ca^{2+} , Zn^{2+} , (100 fold), lysine, cysteine and glucose (60 fold), the results indicated the Cu_2O -CDs/NF/GCE exhibited good selectivity for DA detection.

3.5. Calibration curve

The calibration curve of DA on Cu_2O -CDs/NF/GCE was investigated by differential pulse voltammetry (DPVs) (Fig. 6A). It was found that under optimal experimental conditions the peak current increases with DA concentration, and the oxidation peak current (I_{pa}) has a good linear relationship with DA concentration in the range of 0.05–45.0 μM . As shown in Fig. 6B,

Table 1 Comparison of the analytical performances on different modified electrodes

Electrode	Linear range (μM)	Detection limit (nM)	References
Fe_3O_4 nanoparticle-decorated reduced graphene oxide	0.02–5.80	6.5	32
Nitrogen-doped carbon nanofiber	1.0–200	500	33
N-doped carbon quantum dots	0–1000	1.0	34
Gold nanoparticles/over-oxidized polypyrrole nanotube	0.025–2.5	10	35
Multiwalled carbon nanotubes/electroactive amphiphilic copolymer micelles	0.5–20, 200–1000	200	36
Three-dimensional nitrogen-doped graphene	3.0–100	1.0	37
Gold nanoparticles/tryptophan-functionalized graphene nanocomposite	0.5–411	56	38
Magnetite nanorods/graphene	0.01–100.55	7.0	39
WO_3 nanoparticles	0.1–600	24	19
Cu_2O -CDs/NF	0.05–45.0	1.1	This work

Table 2 Results of determination of DA in human serum ($n = 5$)

Human serum	Spiked (μM)	Found (μM)	Recovery (%)	RSD (%)
Sample 1	5.0	5.33	106.6	3.6
Sample 2	10.0	10.96	109.6	3.2
Sample 3	15.0	14.93	99.5	1.9
Sample 4	20.0	18.88	94.4	2.8
Sample 5	25.0	25.32	101.3	2.4

the linear regression equation was described as $I_{\text{pa}} (\mu\text{A}) = -0.0773C_{\text{DA}} - 2.301$ ($R^2 = 0.998$, $n = 5$), giving a detection limit of 1.1 nM ($S/N = 3$). The detection limit was calculated as 1.1 nM, lower than some previous reports (Table 1) which had been published on RSC Advances and other published in 2015,^{19,32–39} indicating that Cu_2O -CDs/NF/GCE had good sensitivity. The stability of the Cu_2O -CDs/NF/GCE was also examined, the Cu_2O -CDs/NF/GCE was put into a vacuum drying oven at 25 °C, after one week the amperometric responses 99.2% remained against its initial value, 96.5% one month later and 91.3% two months later. The intra- and inter-assay measurements had been also done, and all the experimental errors was under 2%, which indicated the reproducibility for the preparation of electrodes was satisfactory. The excellent characteristics demonstrated the feasibility of its practical application.

3.6. Sample analysis

In order to evaluate the applicability of the proposed method to the determination of DA in real samples, the utility of the developed method was tested by determining DA in human serum (the sample preparation and adequate dilution steps as described earlier).⁴⁰ The results were summarized in Table 2. The recovery rates of the samples ranged between 99.4% and 103.8%, which indicated that the presence of AA, UA and some other substances, such as glucose and sodium chloride did not interfere with the determination of DA. Therefore, the proposed method could be effectively used for the direct determination of DA in real samples.

4. Conclusions

In this study, we have demonstrated a simple combustion method to prepare Cu_2O -CDs with remarkable conductivity by using CDs directly reduce copper hydroxide. Then it was used as working electrode for sensitive and selective determination of DA with the detection limit of 2.2 nM, which are superior to those measured for other metal oxide nanoparticles based working electrode. Due to high yield and efficiency of this method, it may potentially be applied on the scale of industrial production. At the same time, the Cu_2O -CDs/NF/GCE also was applied to the detection of DA content in human serum with satisfactory results, and the biosensor could keep its activity for at least two months.

Acknowledgements

This project was supported by the science and technology foundation of the national general administration of quality supervision in China (no. 2012QK053) and the education bureau of Fujian province of China (no. JB14180).

References

- J. Deng, P. Yu, Y. Wang, L. Yang and L. Mao, *Adv. Mater.*, 2014, **26**, 6933–6943.
- J. Song, J. Li, J. Xu and H. Zeng, *Nano Lett.*, 2014, **14**, 6298–6305.
- J. Deng, P. Yu, Y. Wang and L. Mao, *Anal. Chem.*, 2015, **87**, 3080–3086.
- C. Dong, R. Eldawud, L. M. Sargent, M. L. Kashon, D. Lowry, Y. Rojanasakul and C. Z. Dinu, *Environ. Sci.: Nano*, 2014, **1**, 595–603.
- J. Liu, S. Wagan, M. D. Morris, J. Taylor and R. J. White, *Anal. Chem.*, 2014, **86**, 11417–11424.
- S. M. Majhi, P. Rai, S. Raj, B. S. Chon, K. K. Park and Y. T. Yu, *ACS Appl. Mater. Interfaces*, 2014, **6**, 7491–7497.
- G. Ghadimkhani, N. R. de Tacconi, W. Chanmanee, C. Janaky and K. Rajeshwar, *Chem. Commun.*, 2013, **49**, 1297–1299.
- S. Ni, X. Lv, T. Li, X. Yang and L. Zhang, *Electrochim. Acta*, 2013, **109**, 419–425.
- H. Li, X. Zhang and D. R. MacFarlane, *Adv. Energy Mater.*, 2015, **5**, 1401077.
- F. Xu, M. Deng, G. Li, S. Chen and L. Wang, *Electrochim. Acta*, 2013, **88**, 59–65.
- J. Liu, W. Zhu, S. Yu and X. Yan, *Carbon*, 2014, **79**, 369–379.
- S. Y. Lim, W. Shen and Z. Gao, Carbon quantum dots and their applications, *Chem. Soc. Rev.*, 2015, **44**, 362.
- X. Li, Y. Liu, X. Song, H. Wang, H. Gu and H. Zeng, *Angew. Chem., Int. Ed.*, 2015, **54**, 1759–1764.
- C. Wei, Q. Huang, S. Hu, H. Zhang, W. Zhang, Z. Wang, M. Zhu, P. Dai and L. Huang, *Electrochim. Acta*, 2014, **149**, 237–244.
- C. I. Wang, A. P. Periasamy and H. T. Chang, *Anal. Chem.*, 2013, **85**, 3263–3270.
- Q. Huang, H. Zhang, S. Hu, F. Li, W. Weng, J. Chen, Q. Wang, Y. He, W. Zhang and X. Bao, *Biosens. Bioelectron.*, 2014, **52**, 277–280.
- J. Yang, J. R. Strickler and S. Gunasekaran, *Nanoscale*, 2012, **4**, 4594–4602.
- S. Zheng, Y. Huang, J. Cai and Y. Guo, *Int. J. Electrochem. Sci.*, 2013, **8**, 12296–12307.
- A. C. Anithaa, N. Lavanya, K. Asokan and C. Sekar, *Electrochim. Acta*, 2015, **167**, 294–302.
- S. Hu, Q. Huang, Y. Lin, C. Wei, H. Zhang, W. Zhang, Z. Guo, X. Bao, J. Shi and A. Hao, *Electrochim. Acta*, 2014, **130**, 805–809.
- Y. Lin, P. Yu and L. Mao, *Analyst*, 2015, **140**, 3781–3787.
- R. Devasenathipathy, V. Mani, S. M. Chen, B. Viswanath, V. S. Vasantha and M. Govindasamy, *RSC Adv.*, 2014, **4**, 55900–55907.

- 23 J. Liu, M. D. Morris, F. C. Macazo, L. R. Schoukroun-Barnes and R. J. White, *J. Electrochem. Soc.*, 2014, **161**, H301–H313.
- 24 P. Lu, J. Yu, Y. Lei, S. Lu, C. Wang, D. Liu and Q. Guo, *Sens. Actuators, B*, 2015, **208**, 90–98.
- 25 S. Yuan, W. Chen and S. Hu, *Mater. Sci. Eng., C*, 2005, **25**, 479–485.
- 26 B. Chen, F. Li, S. Li, W. Weng, H. Guo, T. Guo, X. Zhang, Y. Chen, T. Huang, X. Hong, S. You, Y. Lin, K. Zeng and S. Chen, *Nanoscale*, 2013, **5**, 1967–1971.
- 27 C. H. Kuo, C. H. Chen and M. H. Huang, *Adv. Funct. Mater.*, 2007, **17**, 3773–3780.
- 28 M. Bagherzadeh and M. Heydari, *Analyst*, 2013, **138**, 6044–6051.
- 29 E. Laviron, *J. Electroanal. Chem.*, 1979, **101**, 19–28.
- 30 Q. Huang, S. Hu, H. Zhang, J. Chen, Y. He, F. Li, W. Weng, J. Ni, X. Bao and Y. Lin, *Analyst*, 2013, **138**, 5417–5423.
- 31 S. K. Yadav, M. Oyama and R. N. Goyal, *J. Electrochem. Soc.*, 2014, **161**, H41–H46.
- 32 H. Bagheri, A. Afkhami, P. Hashemi and M. Ghanei, *RSC Adv.*, 2015, **5**, 21659–21669.
- 33 J. Sun, L. Li, X. Zhang, D. Liu, S. Lv, D. Zhu, T. Wu and T. You, *RSC Adv.*, 2015, **5**, 11925–11932.
- 34 G. Jiang, T. Jiang, H. Zhou, J. Yao and X. Kong, *RSC Adv.*, 2015, **5**, 9064–9068.
- 35 M. Lin, *RSC Adv.*, 2015, **5**, 9848–9851.
- 36 X. Fei, J. Luo, R. Liu, J. Liu, X. Liu and M. Chen, *RSC Adv.*, 2015, **5**, 18233–18241.
- 37 X. Feng, Y. Zhang, J. Zhou, Y. Li, S. Chen, L. Zhang, Y. Ma, L. Wang and X. Yan, *Nanoscale*, 2015, **7**, 2427–2432.
- 38 Q. Lian, A. Luo, Z. An, Z. Li, Y. Guo, D. Zhang, Z. Xue, X. Zhou and X. Lu, *Appl. Surf. Sci.*, 2015, **349**, 184–189.
- 39 J. Salamon, Y. Sathishkumar, K. Ramachandran, Y. S. Lee, D. J. Yoo, A. R. Kim and G. Gnana kumara, *Biosens. Bioelectron.*, 2015, **64**, 269–276.
- 40 M. Rajamathi, J. S. Melo and T. V. Venkatesha, *Bioelectrochemistry*, 2011, **81**, 104–108.

www.spm.com